

The impact of walking on spinal cord tissue regeneration in patients with paraplegia following spinal cord Injury

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ABSTRACT

The impact of walking in Spinal Cord Injury can promote axonal growth through directed neuroplasticity. The impact of walking in corticospinal tracts, in combination with proprioception, could be the key to neuroregeneration. Furthermore, growth factors such as brain-derived neurotrophic factor (BDNF) and insulin growth factor -1 (IGF-1) play a crucial role not only in the procedure of axonal growth but also in the remyelination.

Many posttraumatic treatment strategies have been evaluated until now, including pharmacological agents aiming to block the development of secondary apoptotic mechanisms of CNS. The same strategies are simultaneously able to promote the regeneration of neuroaxons. Nevertheless, there is insufficient knowledge concerning the hypothesis that gait training could be applied as a potential therapy for neuroprotection following SCI.

The objective of this review is to assess the impact of assisted walking in paraplegia by consolidating evidence regarding: (a) neuroplasticity (b) tissue regeneration.

Key Words: "SCI", "neuroplasticity", "regeneration", "paraplegia", "proprioception", "gait training"

Introduction

According to the International Spinal Cord Society IS-CoS, Spinal cord injury (SCI) is a severe neural trauma that, depending on the severity of the damaged segment, is classified into complete and incomplete. Statistically, it is assessed that 0.0022% of the global population will suffer from a spinal cord injury (SCI), annually [1]. Most of the patients with complete SCI are considered as clinically incomplete due to a few remaining neuron connections based on EMG results [2,3].

Paraplegia refers to the impairment and loss of motor, sensory and/or autonomic function in the thoracic, lumbar or sacral segments [4]. It is a result of severe damage to the spinal cord and the nervous system [5,6]. Recovery following SCI is proved to be perplexed and requires long-term rehabilitation [4]. A hallmark of posttraumatic SCI is neuroplasticity, enabling nervous system to modify and change neural networks to adapt both neuronal structure and function [7-10].

The process of tissue regeneration following SCI

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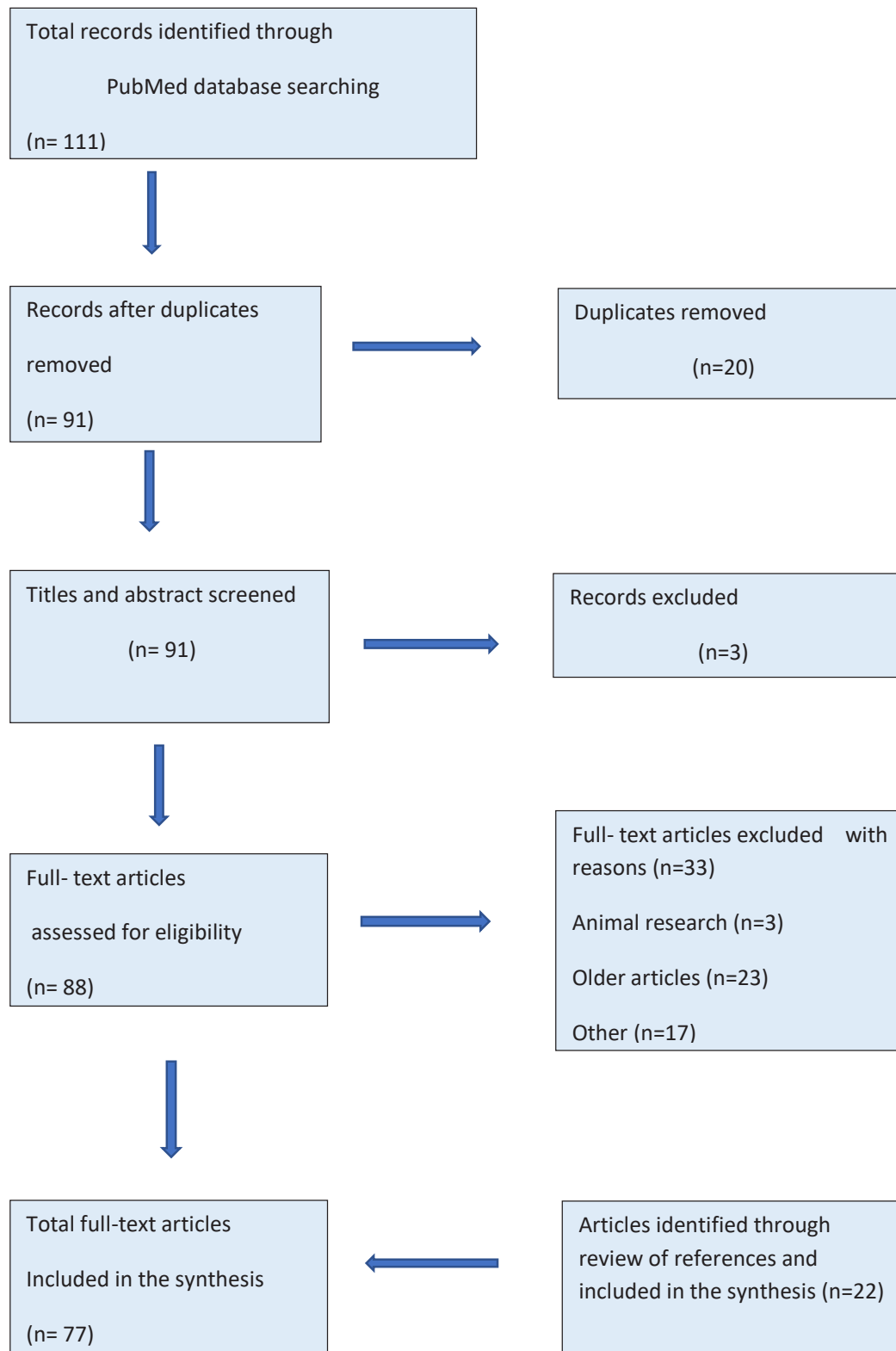


Figure 1: Flowchart of the Article Selection and Review Process

could be differentiated into two phases, the subacute and the chronic phase [5]. The subacute phase refers to some weeks following injury and the chronic to some months-years post-trauma [9]. Walking has been always a goal of the chronic-phase SCI rehabilitation program, concerning locomotion function [11-13]. Nonetheless, there are questions on how walking can affect neuroplasticity and tissue regeneration. This is a potential oversight to assume walking as a therapy.

The main purpose of this review is to provide proof that locomotion training on the treadmill [61], gait-assisted orthosis such as Locomat and wearable exoskeleton robots are capable to foster spinal cord tissue regeneration and promote neuroplasticity through re-learning gait, in individuals with SCI [10]. Although the existing data about this issue is limited and quite perplexing, especially in human subjects, there are some research articles, meta-analysis and systematic reviews that could prove this hypothesis. Successful and functional nerve tissue regeneration should be supported from supraspinal tracts [62], spinal and peripheral inputs to be long-lasting and functional [63].

Pubmed database was reviewed to identify ways that walking can foster spinal cord tissue regeneration and promote neuroplasticity. The key words "SCI", "neuroplasticity", "regeneration", "paraplegia", "proprioception" and "gait training" were used. Search parameters were specific regarding date of publication (from 1995 until 2022) and relevance. The process of identification and the criteria of inclusion-exclusion are described on the following flowchart (Figure 1).

Discussion

The initial electronic database search resulted in a total of 111 articles, of these, 77 were considered for inclusion in this review (Figure 1).

Pathophysiology of Spinal Cord Injury

SCI consist of two major pathophysiological mechanisms, primary and secondary [5,6,14]. The primary refers to the immediate mechanical traumatic damage of spinal cord, leading to demyelination and axotomy [15-17]. Secondary injury involves the presence of free radicals due to the long ischemia and hypoxia [4-6]. Neurotransmission is impaired, lipid peroxidation and calcium influx contribute to apoptosis and axonal de-

myelination [4,14-17]. Finally, scar and cavitation are formed, inhibiting myelin regeneration and limiting axon growth [4,5].

Neuroplasticity of Spinal Cord

Based on the metaplasticity theory, morphology and function of a synapse can change over time [9,18]. Following SCI, the following plasticity includes the neural circuit reconstruction, activation of neurons and nerve conduction [8,19]. The regulation of microenvironment and molecularis aim at neuroprotection of intact axons and gene regulation [20,21]. Specifically, neurotrophic factors such as the brain derived nerve factor-BDNF and insulin growth factor-1 (IGF-1) have been proved to promote adaptive spinal plasticity [20,21,28]. As an adaptive spinal plasticity can be referred as task-dependent plasticity which can be succeed by special forms of training such as stepping [11,19,22].

In patients with paraplegia, stepping may be an effective approach to direct and enhance plasticity however the major question is how [25]. Assisted walking can be defined as a direct-task plasticity specific training of motor function [25]. Hence, the improvement of functional ability of individuals with paraplegia can be translated into enhancement in neuroplasticity. The parameters of these indications are gait speed, intensity, and duration of gait training [25,27]. Literature data support that the improvement of intensity and gait speed are proved to be an indicator of increased plasticity [27].

Research conducted by Leech et al investigated that high-intensive treadmill training in patients with incomplete chronic SCI has an important influence on serum concentration of the brain-derived neurotrophic factor BDNF [28]. The authors demonstrated increased levels of peripheral sBDNF after acute intensive gait training of 11 individuals with SCI [28]. Except from sBDNF, the researchers proved that treadmill training can also affect IGF-1 serum concentration levels [28]. Although there was no correlation between speed and intensity and serum levels of IGF-1, treadmill training increased the levels of IGF-1 following exercise in general [28]. Taking into consideration that insulin-growth factor-1 (IGF-1) is involved in synaptic protein synthesis and interacts with BDNF [20,21], this means that neuroregeneration can be promoted also on a molec-

ular level. Neurotrophic factor BDNF promotes adaptive plasticity which is required for the creation of functional neural connections in SCI [28].

From Tissue Regeneration to Functional Synapsis

The scientific issue of neurogenesis and neuroregeneration in neuroscience has not been completely solved [23,24]. Nowadays, there is limited research data on neural repair following SCI and the mechanisms underneath axon restoration and reconstruction [23]. Nevertheless, the function and efficiency of tissue regeneration mechanisms is still under investigation [5]. Anatomically, there are three broad routes to successfully achieving restitution of functional circuitry [6,18]. The one is the regeneration of damaged axons in long distance, the other is sprouting of lesioned neuron and get connected with the intact ones or alternatively sprouting of undamaged axons and properly joined with the cut neuron [6,18]. The three broad routes alone are inadequate for functional synapsis [18]. Hence, there are some presuppositions that contribute to functional restitution [6,76]. The damaged axons must appropriately guide to the right direction, maintain a long-distance axon growth, circumvent the glial scars, develop mechanisms of remyelination and finally form functional synapses with the intact neurons [18,77].

The remyelination of axons after SCI is an overly complex process [15]. Nevertheless, it is generally accepted that the trophic factor BDNF, Schwann cells and oligodendrocyte precursor cells are involved in remyelinating spared axons, thereby contributing to tissue regeneration [15,16,26]. The growth factor BDNF has been proved to promote adaptive spinal plasticity, thus it can be assumed that there is an interaction between the process of neuroplasticity of spinal neurons and myelin production [28,29]. Furthermore, Schwann cells are well known as the myelinating cells of peripheral nervous system [29,30]. However, they can gain access to spinal axons and assist in forming myelin [30]. This means that peripheral nervous system can influence in a positive way the damaged spinal circuits as far as myelin regeneration is concerned [15]. Eventually, oligodendrocyte precursor cells (OPCs) which originate both from the subventricular zone (SVZ) and locally, migrate to the

SCI lesion site to differentiate into myelin-producing oligodendrocytes [15,31,32].

Axon outgrowth can be sustained only if axonal debris of the damaged neurons can be surpassed. Microglia is the major innate immune cell class in the brain and spinal cord [33]. Using its phagocytosis function, microglia could participate in the maintenance of structural and functional homeostasis of the central nervous system, such as normal myelin turnover and activity-dependent synaptic plasticity [33]. This phagocytosis is significantly up-regulated after injury, as a part of the injury-associated inflammatory responses, to engulf damaged neuronal and axonal debris [33].

Proprioceptive Afferent Promote Neuroplasticity

Proprioceptive feedback incorporated signals from ankle extensors during stance and swing phases of walking [19,34,35]. To begin with, the leg extensor muscles provide load-related afferent information to the spinal cord [36,37]. The activation of these muscles is produced through loading of the sole of the foot during stance phase [37,38]. These activations occur a hip-joint related afferent input, which are appropriate for the initiation of the swing phase [38,39,40]. A study demonstrated that without loading the sole of the foot during the stance phase, no meaningful leg muscle activation occurs in people with complete paraplegia during supporting stepping [41].

Another principal factor that depends on afferent proprioception is the automatism of moving limbs [42]. Spinal automatism is provided during stepping, influencing the coordination of the limbs during locomotion [19,36,42]. In SCI, there is need for training, imitating the automatism and coordination of normal stepping [19,42]. Thus, a basic requirement to induce a locomotion pattern in the thoracolumbar spinal cord can be an assisting training, providing sufficient proprioceptive feedback [36].

Training effects in patients with SCI depend on some number of physiological prerequisites necessary to evoke a pattern of muscle activation like those individuals without SCI to promote adaptive neuroplasticity [19,43,44]. A fundamental factor that is required to trigger a locomotor EMG pattern in SCI patients is an afferent input from load receptors [39,45]. Proprioceptive inflow from leg extensor muscles and mechan-

oreceptors in the sole of the foot, provide load-related afferent information [19,41]. Hip joint-related afferent inputs also play a major role in the generation of a locomotor EMG pattern, in individuals with incomplete spinal cord injury [19,41].

The activation of load receptors and hip-joint related proprioceptive receptors (hip extension) lead to a physiological leg muscle activity pattern during stepping [38]. The research proved that proprioceptive input produced during assisted walking led to targeted leg muscle activation. Electromyographic activity (EMG) of patients with SCI showed great improvement and similarities to normal walking [34,40,46].

Locomotion Rehabilitation Influence Plasticity and Regeneration

It appears that, through gait training, the spinal cord acquires the ability to respond to the imposed patterns of sensory inputs [47,48]. The aim is to retrain the nervous system, stimulating a form of learning that regenerates the surviving circuits and promotes new neuronal connections [22,48]. Stepping, as a spinal learning process, can strengthen the efficacy of neural pathways [47].

Neural excitability within the spinal cord is regulated by motoneurons / interneurons [49]. Khan et al demonstrated that intensive walking training counterpoise the abnormal H-reflex, clonus and stretch reflex following SCI [41,49-52]. Furthermore, the increased impact of corticospinal inputs to interneurons [53] due to training strengthened the inhibitory control of spinal circuits [49]. Additional research demonstrated that gait-assisted training generated and modulated soleus H-reflex [54,66]. Consequently, gait training strengthens spinal descending control and promotes adaptive plasticity [50,51,52].

Cortical and spinal excitability must be increased to activate pyramidal axons and interneurons including spinal intrinsic networks [65,67]. Thomas et al., examine the impact of intensive treadmill training on human spared corticospinal pathways directly [68] and not only based on walking function parameters [11,13]. The authors evaluated patients with chronic SCI and demonstrated that corticospinal tract function was enhanced after intensive daily treadmill training for several months [68]. The researchers

used the system of TMS-evoked MEPs to measure corticospinal excitability, which was quite increased after training. Moreover, the patients were examined again after 2,5 years, and the results were maintained [68]. Therefore, the results of motor evoked potentials (MEP) might be an indication of nerve sprouting on spinal level [57,64].

Spared corticospinal tract (CST)[65] and proprioceptive afferent (PA) axons are both able to sprout after injury and contribute to rewiring spinal circuits, affecting motor recovery [70]. Proprioceptive afferents and descending motor pathways, including the CST, are the two major classes of extrinsic inputs to spinal segmental motor circuits [70,71]. They closely interact with each other during postural control, locomotion, and voluntary movements [70-72]. Both ipsilateral CST and PA axons are very sparse in lamina V to VII of spinal cord and there is minimal overlap between them [70-72]. Taking this fact for granted, it can be a parallel sprouting of proprioceptive afferents and CST axons [27].

Robot- assisted gait training in Paraplegia

Robot-assisted gait training (RAGT) is separated into two categories: the grounded exoskeleton robots (Locomat) [22,34,55] and the wearable exoskeleton robots [76] such as ReWalk [56], Ekso, REX, Indego, and HAL [13,57]. FDA or CE officially approves all [22]. The major difference of these two robot systems is that Locomat is a stationary walking system including a grounded exoskeleton based on a treadmill, whereas the wearable exoskeleton utilizes various environments for gait training [22]. The main purpose of the two exoskeleton robots is to support SCI patients' bodyweight to re-educate walking [58]. Studies proved that RAGT improve neuromuscular relearning in patients with SCI [55,59,60].


Important research with 11 incomplete patients with paraplegia provided supportive evidence for the clinical potential of gait-assisted training with an exoskeleton. Particularly, all participants were trained with the assistance of BWSTT with HAL exoskeleton for 12 weeks [57]. The authors reported that plastic changes appeared in sensorimotor cortical region (S1) after training and long ago after it [57]. Nevertheless, more research is required in this field.

A case study with four patients with tetraplegia evaluated brain plasticity after training with body weight-supported treadmill (BWSTT) and Locomat orthosis [69]. Evidence of great significance was found by the investigators, who demonstrated supraspinal plasticity after 12-week training [69]. The assessment was made with fMRI imaging, proving an increased activation of sensorimotor cortical region (S1,S2) [12,69]. Furthermore, for the participants who achieved functional improvement in over-ground locomotion, the fMRI depicted an enhanced activation of cerebellum [69]. Therefore, it can be assumed that these results may be a consequence of axonal regeneration on spinal level. Consequently, further research should be performed to investigate whether spinal tissue regeneration can be associated with previous results.

Limitations

One limitation of this study is that there was limited exclusive data available on paraplegic patients, since most of research papers also include quadriplegics due to the difficulty of finding subjects for research. Another restriction was the diversity of evaluating techniques used in the studies. For instance, some researchers use fMRI depiction and others measure the concentration of molecules in serum levels.

Conclusion

This review is the first to refer to locomotion rehabilitation as a method to promote nerve tissue regeneration. There are many presuppositions for a successful sprouting and nerve connection. The overall approach of supraspinal, spinal and peripheral tracts can be the key to directed plasticity. 

REFERENCES

1. Rossignol S, Schwab M, Schwartz M, et al. Spinal Cord Injury: Time to move? *Journal of Neuroscience* 31 October 2007; 27(44):11782-92
2. McKinley W, Santos K, Meade M, et al. Incidence and outcomes of spinal cord injury clinical syndromes. *The journal of spinal cord medicine* 2007;30(3): 215-24.
3. Filli L, Schwab ME. Structural and functional reorganization of propriospinal connections promotes functional recovery after spinal cord injury. *Neural Regen Res.* 2015;10(4):509-13.
4. Yang B, Zhang F, Cheng F, et al. Strategies and prospects of effective neural circuit reconstruction after spinal cord injury. *Cell Death Dis* 2020;11:439
5. Ahuja CS, Fehlings M. Concise Review: Bridging the Gap: Novel Neuroregenerative and Neuroprotective Strategies in Spinal Cord Injury. *Stem Cells Transl Med.* 2016; doi: 10.5966/sctm.2015-0381 Epub 2016 Apr 29.
6. Webb AA, Ngan S, Fowler JD. Spinal cord injury I: A synopsis of the basic science. *Can Vet J.* 2010;51(5):485-92.
7. Onifer SM, Smith GM, FouadK. Plasticity after Spinal Cord Injury: Relevance to Recovery and Approaches to Facilitate It. *Neurotherapeutics* 2011;8:283-293.
8. Gassert R, Dietz V. Rehabilitation robots for the treatment of sensorimotor deficits: a neurophysiological perspective. *J NeuroengRehabil.* 2018;15(1):46
9. Grau JW, Huie JR, Lee KH, et al. Metaplasticity and behavior: how training and inflammation affect plastic potential within the spinal cord and recovery after injury. *Front Neural Circuits.* 2014;8:100.
10. De Leon RD, Roy RR, Edgerton VR. Is the recovery of stepping following spinal cord injury mediated by modifying existing neural pathways or by generating new pathways? A perspective. *Physical Therapy* 2001; 81(12):1904-1911.
11. Khan AS, Livingstone DC, Hurd CL, et al. Retraining walking over ground in a powered exoskeleton after spinal cord injury: a prospective cohort study to examine functional gains and neuroplasticity. *J NeuroengRehabil.* 2019;1:145.

12. O'Dell DR, Weber KA, Berliner JC, et al. Mid-sagittal tissue bridges are associated with walking ability in incomplete spinal cord injury: A magnetic resonance imaging case series. *J Spinal Cord Med.* 2020; doi: 10.1080/10790268.2018.1527079 Epub 2018 Oct 22.
13. Zierliacks A, Aach M, Brinkemper A, et al. Rehabilitation of Acute Vs. Chronic Patients with Spinal Cord Injury with a Neurologically Controlled Hybrid Assistive Limb Exoskeleton: Is There a Difference in Outcome? *Front Neurobot.* 2021;15:728327.
14. Donovan J, Kirshblum S. Clinical Trials in Traumatic Spinal Cord Injury. *Neurotherapeutics* 2018;15:654-68.
15. Huntmer-Silveira A, Patil N, Brickner MA, et al. Strategies for Oligodendrocyte and Myelin Repair in Traumatic CNS Injury. *Front. Cell. Neurosci.* 2021;14:619707.
16. Nagappan PG, Chen H, Wang DY. Neuroregeneration and plasticity: a review of the physiological mechanisms for achieving functional recovery postinjury. *Military Med Res* 2020;7:30.
17. Ahuja CS, Nori S, Tetreault L, et al. Traumatic Spinal Cord Injury-Repair and Regeneration. *Neurosurgery.* 2017;80(3):9-22.
18. Dietz V, Fouad K. Restoration of sensorimotor functions after spinal cord injury. *Brain* 2014;137(3): 654-667.
19. Behrman AL, Bowden MG, Nair PM. Neuroplasticity after spinal cord injury and training: an emerging paradigm shift in rehabilitation and walking recovery. *Physical therapy* 2006;86(10):1406-1425.
20. Carro E, Nuñez A, Busiguina S, et al. Circulating insulin-like growth factor I mediate effects of exercise on the brain. *Journal of Neuroscience* 2000;20(8): 2926-2933.
21. Ding Q, Vaynman S, Akhavan M, et al. Insulin-like growth factor I interface with brain-derived neurotrophic factor-mediated synaptic plasticity to modulate aspects of exercise-induced cognitive function. *Neuroscience* 2006; 140(3):823-33.
22. Zhang L, Lin F, Sun L, et al. Comparison of Efficacy of Lokomat and Wearable Exoskeleton-Assisted Gait Training in People with Spinal Cord Injury: A Systematic Review and Network Meta-Analysis. *Front Neurol.* 2022;13:772660.
23. Zheng Y, Mao YR, Yuan TF, et al. Multimodal treatment for spinal cord injury: a sword of neuroregeneration upon neuromodulation. *Neural Regen Res.* 2020;15(8):1437-50.
24. Shah M, Peterson C, Yilmaz E, et al. Current advancements in the management of spinal cord injury: A comprehensive review of literature. *Surg Neurol Int.* 2020;11:2.
25. Stevenson AJ, Mrachacz-Kersting N, van Asseldonk E, et al. Spinal plasticity in robot-mediated therapy for the lower limbs. *J Neuroeng Rehabil.* 2015;12:81.
26. Hopf A, Schaefer DJ, Kalbermatten DF, et al. Schwann Cell-Like Cells: Origin and Usability for Repair and Regeneration of the Peripheral and Central Nervous System. *Cells.* 2020; 9(9):1990.
27. Wirz M, Zemon DH, Rupp R, et al. Effectiveness of automated locomotor training in patients with chronic incomplete spinal cord injury: a multicenter trial. *Archives of physical medicine and rehabilitation* 2005; 86(4):672-680.
28. Leech KA, Hornby TG. High-Intensity Locomotor Exercise Increases Brain-Derived Neurotrophic Factor in Individuals with Incomplete Spinal Cord Injury. *J Neurotrauma.* 2017;doi: 10.1089/neu.2016.4532 Epub 2017 Jan 18.
29. Kwon BK, Fisher CG, Dvorak MF, et al. Strategies to Promote Neural Repair and Regeneration After Spinal Cord Injury. *Spine* 2005;30(17): 3-13
30. Pearse DD, Pereira FC, Marcillo AE, et al. cAMP and Schwann cells promote axonal growth and functional recovery after spinal cord injury. *Nat Med.* 2004;doi: 10.1038/nm1056 Epub 2004 May 23.
31. Watson RA, Yeung TM. What is the potential

- of oligodendrocyte progenitor cells to successfully treat human spinal cord injury? *BMC Neurol* 2011;11:113.
32. Li N, Leung GK. Oligodendrocyte Precursor Cells in Spinal Cord Injury: A Review and Update. *Biomed Res Int*. 2015; doi: 10.1155/2015/235195 Epub 2015 Sep 27.
 33. Jiang YQ, Armada K, Martin JH. Neuronal activity and microglial activation support corticospinal tract and proprioceptive afferent sprouting in spinal circuits after a corticospinal system lesion. *Exp Neurol*. 2019; doi: 10.1016/j.expneurol.2019.113015 Epub 2019 Jul 18.
 34. Dietz V, Müller R, Colombo G. Locomotor activity in spinal man: significance of afferent input from joint and load receptors. *Brain* 2002; 125(12):2626-34.
 35. Shin JC, Jeon HR, Kim D, et al. Effects on the Motor Function, Proprioception, Balance, and Gait Ability of the End-Effector Robot-Assisted Gait Training for Spinal Cord Injury Patients. *Brain Sciences*. 2021;11(10):1281.
 36. Zehr EP. Neural control of rhythmic human movement: the common core hypothesis. *Exerc Sport Sci Rev*. 2005;33(1):54-60.
 37. Dietz V. Proprioception and locomotor disorders. *Nature Reviews Neuroscience* 2002; 3(10):781-790.
 38. Qaiser T, Eginyan G, Chan F, et al. The sensorimotor effects of a lower limb proprioception training intervention in individuals with a spinal cord injury. *J Neurophysiol*. 2019; doi:10.1152/jn.00842.2018 Epub 2019 Oct 30.
 39. Dietz V, Harkema SJ. Locomotor activity in spinal cord-injured persons. *Journal of Applied Physiology* 2004;96(5):1954-60.
 40. Dietz V, Colombo G, Jensen L, et al. Locomotor capacity of spinal cord in paraplegic patients. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society* 1995;37(5):574-82.
 41. Smith AC, Mummidisetty CK, Rymer WZ, et al. Locomotor training alters the behavior of flexor reflexes during walking in human spinal cord injury. *J Neurophysiol*. 2014; doi:10.1152/jn.00308.2014 Epub 2014 Aug 13.
 42. Clark DJ. Automaticity of walking: functional significance, mechanisms, measurement and rehabilitation strategies. *Front Hum Neurosci*. 2015;9:246.
 43. Hubli M, Dietz V. The physiological basis of neurorehabilitation--locomotor training after spinal cord injury. *J NeuroengRehabil*. 2013;10:5.
 44. Ivanenko YP, Poppele RE, Lacquaniti F. Five basic muscle activation patterns account for muscle activity during human locomotion. *J Physiol*. 2004; doi: 10.1113/jphysiol.2003.057174 Epub 2004 Jan 14.
 45. Hayes SC, White M, Wilcox CRJ, et al. Biomechanical differences between able-bodied and spinal cord injured individuals walking in an overground robotic exoskeleton. *PLoS One*. 2022 Jan 27;17(1):e0262915.
 46. Dietz V, Müller R. Degradation of neuronal function following a spinal cord injury: mechanisms and countermeasures. *Brain* 2004; 127(10):2221-31.
 47. Grau JW, Baine RE, Bean PA, et al. Learning to promote recovery after spinal cord injury. *Exp Neurol*. 2020; doi: 10.1016/j.expneurol.2020.113334 Epub 2020 Apr 28.
 48. Smith GM, Falone AE, Frank E. Sensory axon regeneration: rebuilding functional connections in the spinal cord. *Trends Neurosci*. 2012; doi:10.1016/j.tins.2011.10.006 Epub 2011 Nov 30.
 49. Khan AS, Patrick SK, Roy FD, et al. Training-Specific Neural Plasticity in Spinal Reflexes after Incomplete Spinal Cord Injury. *Neural Plast*. 2016; doi: 10.1155/2016/6718763 Epub 2016 Sep 20.
 50. Smith AC, Knikou M. A Review on Locomotor Training after Spinal Cord Injury: Reorganization of Spinal Neuronal Circuits and Recovery of Motor Function. *Neural Plast*. 2016; doi: 10.1155/2016/1216258 Epub 2016 May 11.
 51. Knikou M, Mummidisetty CK. Locomotor training improves premotoneuronal con-

- control after chronic spinal cord injury. *J Neurophysiol.* 2014;111(11):2264-75. doi: 10.1152/jn.00871.2013 Epub 2014 Mar 5.
52. Smith AC, Rymer WZ, Knikou M. Locomotor training modifies soleus monosynaptic motoneuron responses in human spinal cord injury. *Exp Brain Res.* 2015; doi: 10.1007/s00221-014-4094-7 Epub 2014 Sep 10.
53. Zholudeva LV, Abaira VE, Satkunendrarah K, et al. Spinal Interneurons as Gatekeepers to Neuroplasticity after Injury or Disease. *J Neurosci.* 2021; doi: 10.1523/Jneurosci.1654-20.2020 Epub 2021 Jan 20.
54. Knikou M, Smith AC, Mummidisetty CK. Locomotor training improves reciprocal and nonreciprocal inhibitory control of soleus motoneurons in human spinal cord injury. *Journal of Neurophysiology* 2015; 113(7):2447-60.
55. Mirbagheri MM, Tsao C, Pelosin E, et al. Therapeutic effects of robotic-assisted locomotor training on neuromuscular properties. In *9th International Conference on Rehabilitation Robotics*, 2005. ICORR 2005. (pp. 561-564). IEEE.
56. Alamro RA, Chisholm AE, Williams AMM, et al. Overground walking with a robotic exoskeleton elicits trunk muscle activity in people with high-thoracic motor-complete spinal cord injury. *J NeuroengRehabil.* 2018;15(1):109.
57. Sczesny-Kaiser M, Höffken O, Aach M, et al. HAL® exoskeleton training improves walking parameters and normalizes cortical excitability in primary somatosensory cortex in spinal cord injury patients. *J NeuroengRehabil.* 2015;12:68.
58. Calabrò RS, Naro A, Leo A, et al. Usefulness of robotic gait training plus neuromodulation in chronic spinal cord injury: a case report. *J Spinal Cord Med.* 2017; doi: 10.1080/10790268.2016.1153275 Epub 2016 Mar 4.
59. Shin JC, Kim JY, Park HK, et al. Effect of robotic-assisted gait training in patients with incomplete spinal cord injury. *Ann Rehabil Med.* 2014; doi: 10.5535/arm.2014.38.6.719 Epub 2014 Dec 24.
60. Nam KY, Kim HJ, Kwon BS, et al. Robot-assisted gait training (Lokomat) improves walking function and activity in people with spinal cord injury: a systematic review. *J NeuroengRehabil.* 2017; doi: 10.1186/s12984-017-0232-3.
61. Dobkin BH, Apple D, Barbeau H, et al. Methods for a Randomized Trial of Weight-Supported Treadmill Training Versus Conventional Training for Walking During Inpatient Rehabilitation after Incomplete Traumatic Spinal Cord Injury. *Neurorehabilitation and Neural Repair.* 2003; 17(3):153-67.
62. Field-Fote EC, Yang JF, Basso DM, et al. Supraspinal Control Predicts Locomotor Function and Forecasts Responsiveness to Training after Spinal Cord Injury. *J Neurotrauma.* 2017; doi: 10.1089/neu.2016.4565 Epub 2016 Dec 20.
63. Barthélemy D, Knudsen H, Willerslev-Olsen M, et al. Functional implications of corticospinal tract impairment on gait after spinal cord injury. *Spinal Cord* 2013; 51: 852-56.
64. Knikou M. Plasticity of corticospinal neural control after locomotor training in human spinal cord injury. *Neural Plast.* 2012; doi: 10.1155/2012/254948 Epub 2012 Jun 4.
65. Oudega M, Perez MA. Corticospinal reorganization after spinal cord injury. *J Physiol.* 2012; doi: 10.1113/jphysiol.2012.233189 Epub 2012 May 14.
66. Querry RG, Pacheco F, Annaswamy T, et al. Synchronous stimulation and monitoring of soleus H reflex during robotic body weight-supported ambulation in subjects with spinal cord injury. *Journal of Rehabilitation Research & Development* 2008;45(1).
67. Blicher JU, Nielsen JF. Cortical and spinal excitability changes after robotic gait training in healthy participants. *Neurorehabilitation and neural repair* 2009; 23(2):143-49.
68. Thomas SL, Gorassini MA. Increases in corticospinal tract function by treadmill training after incomplete spinal cord injury. *Journal of neurophysiology* 2005; 94(4):2844-55.
69. Winchester P, McColl R, Querry R, et al.

Changes in supraspinal activation patterns following robotic locomotor therapy in motor-incomplete spinal cord injury. *Neurorehabilitation and neural repair* 2005; 19(4):313-24.

70. Blecher R, Heinemann-Yerushalmi L, Assaraf E, et al. New functions for the proprioceptive system in skeletal biology. *Philos Trans R Soc Lond B Biol Sci.* 2018; 373(1759):20170327.
71. MacKinnon CD. Sensorimotor anatomy of gait, balance, and falls. *Handb Clin Neurol.* 2018; 159:3-26.
72. Takakusaki K. Functional Neuroanatomy for Posture and Gait Control. *J Mov Disord.* 2017;10(1):1-17.
73. Hillier S, Immink M, Thewlis D. Assessing Proprioception: A Systematic Review of Possibilities. *Neurorehabilitation and Neural Repair.* 2015; 29(10):933-49.
74. Domingo A, Lam T. Reliability and validity of using the Lokomat to assess lower limb joint position sense in people with incomplete spinal cord injury. *J NeuroengRehabil.* 2014;11:167.
75. Angeli CA, Edgerton VR, Gerasimenko YP, et al. Altering spinal cord excitability enables voluntary movements after chronic complete paralysis in humans. *Brain.* 2014 doi: 10.1093/brain/awu038 Epub 2014 Apr 8.
76. Dietz V, Curt A. Neurological aspects of spinal cord repair: promises and challenges. *Lancet Neurol.* 2006;5(8):688-94.
77. Fink KL, Cafferty WB. Reorganization of Intact Descending Motor Circuits to Replace Lost Connections after Injury. *Neurotherapeutics.* 2016; 13(2):370-81.

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CITATION

Mileti A, Vasiliadis E. The impact of walking on spinal cord tissue regeneration in patients with paraplegia following spinal cord Injury. *Acta Orthop Trauma Hell* 2023; 74(1): 93-102.