

Infections of the spine: Current concepts and a literature review

Gavriil P., Sioutis S., Bekos A., Gerasimides P., Georgoulis J., Soultanis K., Mavrogenis A.F., Sapkas G.
*First Department of Orthopaedics, National and Kapodistrian University of Athens, School of Medicine, ATTIKON
University Hospital, Athens, Greece*

ABSTRACT

Infections of the spine comprise a wide spectrum of different clinical manifestations depending on the exact anatomical structure involved. Spinal infections pose an essential health problem, the treatment of which requires a multidisciplinary approach. Diagnosis is based on clinical symptoms, radiologic evidence, laboratory tests and biopsy. The most common pathogens are bacteria; most of which spread hematogenously. Current treatment involves a combination of antibiotic agents. Sometimes, surgery is required to eradicate the infection or to treat its complications. In all cases, thorough and repetitive clinical examination and laboratory tests are of paramount importance for optimal outcomes.

KEY WORDS: Spine Infections, Spondylitis, Spondylodiscitis, Pathogenesis, Clinical Presentation, Back pain

1. Introduction

Infections of the spine and their various clinical manifestations consist a group of challenging medical conditions which necessitate a team of specialists for optimal diagnosis, treatment and recovery. The responsible pathogens are usually bacteria, however, fungi and even parasites can be encountered. Spinal infections can be classified as pyogenic (bacterial), granulomatous (tuberculosis or fungal) or parasitic (Echinococcosis).[1] Alternatively, an anatomical classification can be used. [2]. Depending on the route of spread of the pathogens, spinal infections can be divided in those that spread hematogenously, from adjacent tissues, or through direct inoculation. This is a review of the literature regarding infections of the spine. We also describe and summarize the epidemiology, pathogenesis, clinical manifestation, diagnosis

and management of spinal infections.

2. Epidemiology

Spinal infections are relatively rare with an estimated incidence around 22 cases per million per year. [3] Vertebral osteomyelitis is responsible for about 0.15% to 5% of all osteomyelitis cases.[4] Despite being a rare entity, vertebral osteomyelitis is the most frequent form of osteomyelitis spreading hematogenously in older patients. [5]

The most commonly diagnosed spinal infection is primary pyogenic spondylodiscitis [2],[6]. The causative pathogens are Gram positive bacteria especially *Staphylococcus Aureus*. [7] The disease has a male: female ratio of 1.5.[3],[8] It usually affects people in their 50s or 60s.[9] An exception is younger intravenous drug users.[10] Prior to the use of antibiotics, spondy-

CORRESPONDING
AUTHOR,
GUARANTOR

Mavrogenis A.F., Associate Professor, First Department of Orthopaedics, National and Kapodistrian University of Athens, School of Medicine, ATTIKON University Hospital, Athens, Greece, Mob.: 697 222 60 96
E-mail: afm@otenet.gr, andreasfmavrogenis@yahoo.gr

lodylitis had a mortality ratio of 25–71%. The current rate is 2–12% [11]

The spine can be extensively affected with multifocal or adjoining lesions (common in TB osteomyelitis) or present as an isolated site of infection as in pyogenic cases. [12] The most common region affected is the lumbar spine followed by the thoracic spine. [4], [6] A distinct entity, tuberculous spondylodiscitis has predilection for the thoracolumbar region. [13] Sacral osteomyelitis has been described, usually as a complication of infected pressure ulcers in bedridden patients. [14] The infection may expand posteriorly forming epidural or subdural abscesses, or laterally, forming most commonly psoas abscesses. [15] Facet involvement has been described as septic facet joint arthritis. [16]

In terms of epidemiology, certain risk factors predispose to spinal infection; immunocompromised in particular are in great danger. [5] Another category, intravenous drug addicts incur high likelihood of infection from repetitive injections. [10] Likewise, people with common clinical conditions like diabetes, malignancy, renal or hepatic failure sustain a higher risk for spinal infection. [17], [18] A distinct category of patients with increased likelihood for regional infection are those who had spinal surgery and those with orthopedic or other implants. [19] Moreover, immigrants from third world countries, inmates, and those of low socioeconomic level are exceptionally vulnerable. [20]

3. Pathogenesis

There are two possible routes of dissemination: the hematogenous and the non-hematogenous; the latter is further divided to direct inoculation and contiguous spread. In hematogenous spread bacteria due to simple events like tooth brushing related microtrauma or more serious, like urinary tract infections circulate in the bloodstream. [21] A common source of bacteremia are various kinds of medical implants. Hematogenous spread allows bacterial seeding the metaphyseal and cartilaginous end-plates and afterwards into the adjacent tissue. [22] The characteristic vascular anatomy and physiology of the region provides the appropriate circumstances (slow blood flow, lack of valves) for pathogen adherence and proliferation. The hematogenous route is the most common route of dissemination and perfectly describes the pathogenesis of pyogenic

spondylodiscitis. Once microorganisms enter the vascular arcades in the metaphysis, the infection spreads. The disc is destroyed by bacterial enzymes. [23] Tuberculous infection stems from Batson's paravertebral venous plexus. Tuberculous spondylitis characteristically encompasses early obliteration of the anteroinferior part of vertebral bodies and may then expand beneath, involving the anterosuperior aspect of the inferior vertebra. [12] However, tuberculous spondylitis does not destroy the disc until late disease. [24].

There are two additional, less frequent, ways of pathogen dissemination in spinal infection. The first is direct inoculation, commonly due to regional trauma or recent surgery in the spine or surrounding tissue. [25], [26] The second is contiguous spread from adjacent foci as the aorta, the esophagus or the bowel. [27]

Children and adults manifest differences regarding pathogenesis. In children, the spread of infection is rapid, because vessels supply both the end plates and the intervertebral discs, whereas in adults, intraosseous arteries are end-arteries; septic emboli may occlude the circulation, resulting in broad destruction. [28]

4. Clinical presentation

Awareness of the clinical presentation is crucial in the recognition of spinal infection. [29] Nonetheless, this can be particularly difficult due to the non-specific, and often mild symptoms of spondylodiscitis, especially in early disease. Thus, initial diagnosis delays more than three months after development of the first symptoms in about 50% of the patients. [30]

Idiopathic back or neck pain has often been described as the predominant symptom. [31] Paravertebral muscle tenderness and spasm, and limitation of spine movement represent the predominant signs in spondylodiscitis. [32] Pain should be differentiated from the common back pain. This can be achieved by looking for concomitant "red flags", for instance fever, malaise, neurological deficits, and persistent symptoms with minimum or no improvement. However, fever is rarely present in patients with mycobacterial, brucella, or fungal spondylodiscitis and may be absent in patients taking analgesics. [33]

Clinical examination is necessary and can be very helpful. Inspection of the patient can detect the cause

TABLE 1.

Table 1: Parenteral Antimicrobial Treatment of Common Microorganisms Causing Native Vertebral Osteomyelitis (Ryang, Y.-M., Akbar, M., 2020.)

Microbiology [77], [78]	Incidence (%)	Route of infection
Staphylococcus aureus	20–84	Most common pathogen; 1.7–6% of bloodstream infections complicated by VO
Coagulase-negative staphylococci	5–16	Device-related bacteraemia or direct inoculation in post-operative infections
Streptococci and enterococci	5–20	Haematogenous spread. Associated with infective endocarditis in 26%
Enterobacteriaceae	7–33	Haematogenous spread from urinary tract infections in older population. Commonly Escherichia coli, Proteus, Klebsiella, Enterobacter spp
Anaerobes	<4	Contiguous spread from pelvic or intra-abdominal foci. Cutibacterium acnes direct inoculation from implants
Polymicrobial	<10	Contiguous spread

(scars due to trauma or previous operations). Paravertebral tenderness and masses (muscle spasm or rarely abscess formation) may be palpated. [34]

The role of neurologic examination is crucial because it can unveil neurologic deficits. In such cases, common findings are muscle weakness, sensory impairment or loss and sphincters incompetence.[54]

5. Diagnosis

Any delay in diagnosis increases the risk for abscess formation and confer increased morbidity and mortality.[29] Co-existing medical conditions, previous surgeries and drug use can raise the suspicion for spinal infection or elucidate the primary cause. [11],[18]

Laboratory work up includes White Blood Cells count (WBC), Erythrocyte Sedimentation Rate (ESR) and C - reactive protein (CRP). WBC is slightly elevated or normal in about half the patients with spondylodiscitis, thus is relatively nonspecific. ESR is a more sensitive inflammatory marker, found elevated in > 90% of patients.[36] CRP seems to be the most important blood test, being very sensitive and normalizing in response to treatment.[35] However, these markers remain relatively nonspecific.[37] Blood cultures should be part of routine laboratory evaluation. However, cultures often fail to identify a specific pathogen. [38] Quantification of interferon-gamma (IFN- γ) based

tests for tuberculous infection detection or serologic tests for Brucella can be utilized in patients from endemic areas.[39]

The next step is the use of radiologic modalities. Even though radiographs have low specificity, they remain a valuable, low-cost, diagnostic tool with high sensitivity.[40] Radiographic signs suggesting spondylodiscitis are narrowing of disc space, loss of definition and irregularity of the vertebral endplate. Pedicle, lamina and spinous process involvement is rare in pyogenic spondylodiscitis and should alert for tuberculous infection. [41] Destruction of intervertebral disc is indicative of pyogenic infection.[4], [42]

MR imaging is the modality of choice with 96% sensitivity, and 94% specificity.[43],[44],[45] MRI offers details about paravertebral soft tissue involvement, abscess formation, nerve root and spinal compression. Although gadolinium-enhanced MRI scans are highly sensitive and specific they often overestimate the presence and extent of infection. [46]

Computerized Tomography (CT) can be utilized whenever MRI is contraindicated. Indicative findings of vertebral infection are end-plate erosion, paravertebral fat reduction, disc hypodensity and bone necrosis or pathological calcification. [37], [42]

Technetium or leucocyte labelled bone scintigraphy, although relatively sensitive (90%), has low specificity,

TABLE 2.

Parenteral Antimicrobial Treatment of Common Microorganisms Causing Native Vertebral Osteomyelitis (Barberi et al., 2015)

Microorganism	First Choice ^a	Alternatives ^a	Comments ^b
Staphylococci, oxacillin susceptible	Nafcillin ^c sodium or oxacillin 1.5–2 g IV q4–6 h or continuous infusion or Cefazolin 1–2 g IV q8 h or Ceftriaxone 2 g IV q24 h	Vancomycin IV 15–20 mg/kg q12 h ^d or daptomycin 6–8 mg/kg IV q24 h or linezolid 600 mg PO/IV q12 h or levofloxacin 500–750 mg PO q24 h and rifampin PO 600 mg daily [86] or clindamycin IV 600–900 mg q8 h	6 wk duration
Staphylococci, oxacillin resistant [87]	Vancomycin IV 15–20 mg/kg q12 h (consider loading dose, monitor serum levels)	Daptomycin 6–8 mg/kg IV q24 h or linezolid 600 mg PO/IV q12 h or levofloxacin PO 500–750 mg PO q24 h and rifampin PO 600 mg daily [86]	6 wk duration
Enterococcus species, penicillin susceptible	Penicillin G 20–24 million units IV q24 h continuously or in 6 divided doses; or ampicillin sodium 12 g IV q24 h continuously or in 6 divided doses	Vancomycin 15–20 mg/kg IV q12 h (consider loading dose, monitor serum levels) or daptomycin 6 mg/kg IV q24 h or linezolid 600 mg PO or IV q12 h	Recommend the addition of 4–6 wk of aminoglycoside therapy in patients with infective endocarditis. In patients with BSI, physicians may opt for a shorter duration of therapy. Optional for other patients [88], [89]. Vancomycin should be used only in case of penicillin allergy.
Enterococcus species, penicillin resistant^e	Vancomycin IV 15–20 mg/kg q12 h (consider loading dose, monitor serum levels)	Daptomycin 6 mg/kg IV q24 h or linezolid 600 mg PO or IV q12 h	Recommend the addition of 4–6 wk of aminoglycoside therapy in patients with infective endocarditis. In patients with BSI, physicians may opt for a shorter duration of aminoglycoside. The additional of aminoglycoside is optional for other patients [88], [89]
Pseudomonas aeruginosa	Cefepime 2 g IV q8–12 h or meropenem 1 g IV q8 h or doripenem 500 mg IV q8 h	Ciprofloxacin 750 mg PO q12 h (or 400 mg IV q8 h) or aztreonam 2 g IV q8 h for severe penicillin allergy and quinolone-resistant strains or ceftazidime 2 g IV q8 h	6 wk duration Double coverage may be considered (ie, β -lactam and ciprofloxacin or β -lactam and an aminoglycoside).

Enterobacteriaceae	Cefepime 2 g IV q12 h or ertapenem 1 g IV q24 h	Ciprofloxacin 500–750 mg PO q12 h or 400 mg IV q12 hours	6 wk duration
β-hemolytic streptococci	Penicillin G 20–24 million units IV q24 h continuously or in 6 divided doses or ceftriaxone 2 g IV q24 h	Vancomycin IV 15–20 mg/kg q12 h (consider loading dose, monitor serum levels)	6 wk duration Vancomycin only in case of allergy.
Propionibacterium acnes	Penicillin G 20 million units IV q24 h continuously or in 6 divided doses or ceftriaxone 2 g IV q24 h	Clindamycin 600–900 mg IV q8 h or vancomycin IV 15–20 mg/ kg q12 h (consider loading dose, monitor serum levels)	6 wk duration Vancomycin only in case of allergy.
Salmonella species	Ciprofloxacin PO 500 mg q12 h or IV 400 mg q12 h	Ceftriaxone 2 g IV q24 h (if nalidixic acid resistant)	6–8 wk duration

Abbreviations: BSI, bloodstream infection; IV, intravenous; PO, take orally; q, every.

^a Antimicrobial dosage needs to be adjusted based on patients' renal and hepatic function. Antimicrobials should be chosen based on in vitro susceptibility as well as patient allergies, intolerances, and potential drug interactions or contraindications to a specific antimicrobial.

^b Recommend Infectious Diseases Society of America guidelines for monitoring of antimicrobial toxicity and levels [136]

^c Flucloxacillin may be used in Europe.

^d Vancomycin should be restricted to patients with type I or documented delayed allergy to β-lactams.

^e Daptomycin, linezolid, or Synercid may be used for vancomycin-resistant enterococci.

thus it is not routinely used. A plethora of novel nuclear imaging modalities exist such as ¹¹¹In, Gallium spine scan and streptidion scintigraphy. These modalities are very sensitive and specific, however, the requirement for specialized facilities and personnel, limits their role.[47],[48],[49] Fluorine-18 (F-18) fluorodeoxyglucose-positron emission tomography (FDG-PET) has shown promising results for both acute and chronic infection, being particularly useful in patients with metallic implants because FDG uptake is not hampered by metallic artifacts.[50], [51]

When blood cultures fail to identify a pathogen, biopsy is considered; open or percutaneous. While open biopsy is a last resort option, percutaneous biopsy is routinely executed.[52],[53],[54] In addition to bacterial cultures, mycobacterial, brucella and fungal cultures should be obtained.[55], [56] If the results are inconclusive, a second CT-guided needle biopsy may be performed before open biopsy is finally required.[57] In either case PCR should be used. Molecular diagnostic tools have improved the yield of microbiologic diagnosis via tissue biopsy.[58],[59] Use of antimicrobial agents before biopsy remains a highly debatable topic. We recommend adhering to the classical approach and withholding initiation of treatment when this is

feasible.[60],[61],[62] In patients with neurologic compromise or hemodynamic instability, we recommend immediate surgical intervention plus empiric antimicrobial therapy.[63]

6. Differential diagnosis

Diagnosis of spinal infection based on clinical signs and symptoms is very challenging. Initial differential diagnosis consists of common causes of back and neck pain such as trauma, disc herniation, osteoporosis, rheumatic diseases and pathologic conditions as malignancies.

A distinction between mechanical causes and pathologic conditions can be presumed clinically. Back pain that resolves with bed rest and limitation of physical activity points towards mechanical causes. On the other hand, pain of insidious onset with evolving neurologic deficits, prolonged pain, aggravating at night or with rest and accompanied by other general signs and symptoms should raise awareness for pathologic conditions. Imaging and biochemical, microbiological and histopathological evaluation should be considered.

7. Microbiology

Epidemiology of the causative pathogens of spinal in-

fections varies. Vertebral osteomyelitis can be polymicrobial, albeit usually one pathogen is responsible.[23] The infectious microorganisms are bacteria, fungi or rarely parasites; bacteria remain the predominant cause of the disease. Specifically, gram positive cocci are responsible for the most common type of spinal infection: pyogenic vertebral osteomyelitis, whereas in the past, tuberculous osteomyelitis was the commonest.[64] Although uncommon in Western world nowadays, TB remains an important cause of spinal infection in endemic countries. Patients with tuberculous spinal infection, not coming from an endemic area typically are immunocompromised or elders, possibly reflecting reactivation of a latent infection.[65] In extreme cases, spondylodiscitis is a complication of intravesical BCG (bacillus Calmette-Guerin) instillation in people treated for bladder cancer.[66] *Staphylococcus aureus* is the most common isolated bacterium, responsible for 20% to 84% of all spinal infections.[7],[67] *Staphylococcus lugdunensis* has been associated with deep-seated infections and may mimic *S. aureus*. [68] *Staph. Epidermitis*, related with iatrogenic or periprosthetic infection, has been linked with cases of spondylodiscitis.[69] Streptococci and Enterococci related spinal infections represent 5% to 20% of cases.[40] Enterobacteriae species follow with about the same incidence (7-33%). They are strongly related with concomitant urinary tract or gastrointestinal infections. *Salmonella* species have been linked with vertebral osteomyelitis in children, particularly those with sickle cell disease[70]. Another causative pathogen for spinal infection in children is *Kingella Kingae*, however, it is not routinely isolated. [71] *Pseudomonas aeruginosa*, a rare pathogen, is found in 0% to 6% overall positive bacterial cultures.[72],[73] IV drug abusers are more likely to be infected with *Pseudomonas*. [74] *Cutibacterium Acnes* has been implicated as causative pathogen for spinal infection, despite previously considered iatrogenic contaminant. Implant associated contamination during orthopedic surgeries is another way of seeding.[75],[76]

Brucella species should be considered in endemic areas, accounting for 30% of spinal infections.[3], [79],[80] Fungal spinal infection is rare and can occur in patients in endemic areas or certain host risk factors such as immunocompromised (*Aspergillus*), intravenous drug users or indwelling intravenous catheters (*Candida*,



Figure 1. Pyogenic spondylitis of the L3 and L4 vertebrae after facet joint injections successfully treated with debridement and antibiotics.

Aspergillus). [81],[82] Parasitic infections are extremely rare globally but common in endemic areas. Specifically, spinal echinococcosis, due to *Echinococcus granulosus*, is found in sheep breeding areas of the Eastern and Southern countries of Mediterranean sheep breeding. Thus, awareness and clinical suspicion is necessary in patients coming from these regions.[83]

8. Conservative treatment

The next step is appropriate therapeutic management. Conservative treatment is the treatment of choice in uncomplicated spondylodiscitis and those who are not candidates for surgical operation. Conservative treatment involves antibiotics, analgesics, special spinal braces, physiotherapy and immobilization. The goal is pain suppression, infection eradication and ensuring the stability of the vertebral column.[84]

Regarding immobilization, usually a period of bed rest (1-2 weeks) followed by a period of patient ambulation using special rigid braces is applied. Prolonged bed rest (up to six weeks) is associated with complications such as thrombi and pulmonary emboli, thus should be applied only when necessary. Generally, early ambula-

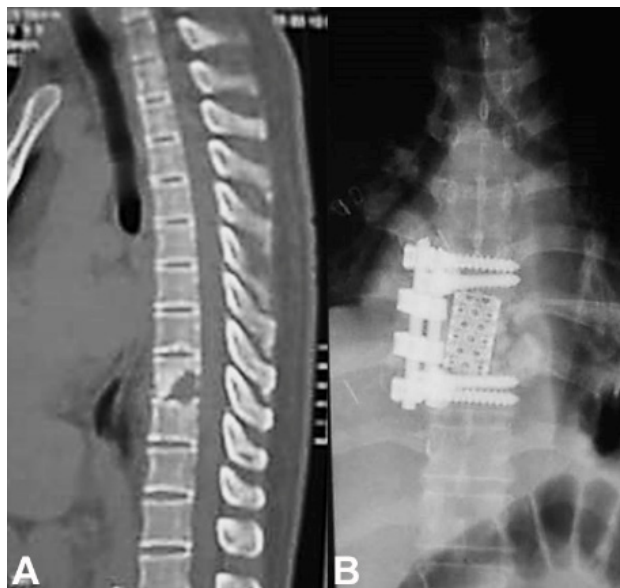


Figure 2. (A) TBC spondylitis of the T9 vertebra (B) successfully treated with vertebrectomy and fusion, and antituberculous medication for 12 months.

tion with spinal braces should be encouraged. [85]

Antibiotics are used invariably in the clinical management of patients with spinal infection. Generally, in patients with hemodynamic instability, progressive or severe neurologic symptoms empirical antimicrobial therapy is initiated, whereas in stable patients selective antimicrobial therapy based on the specific pathogen and susceptibility tests is applied.[61] According to IDSA 2015 guidelines, empiric regimen should cover for staphylococci, including MRSA, streptococci, and gram-negative bacilli. Such regimens include a combination of vancomycin and a third- or fourth-generation cephalosporin. In case of allergy or intolerance, daptomycin and quinolone are reasonable alternatives.[23] Common therapeutic regimen are shown in the following table:

Treatment of spinal tuberculosis necessitates a complicated combination of antimicrobial agents.[91] A commonly used protocol constitutes of isoniazid, rifampicin, ethambutol, and pyrazinamide.[92] *Brucella* spondylodiscitis is treated with a combination of either streptomycin plus doxycycline or rifampin plus doxycycline.[11] Management of patients with fungal spinal infection involves a variety of drugs; azoles and amphotericin B are the most common choices.[93],[94]

Prolonged antibiotic treatment is recommended due to the limited bone penetration of most antimicrobials. [95],[96] Nevertheless, the optimal duration remains a debatable topic with most studies suggesting a 6-8 week regimen.[97] Accordingly, the 2015 IDSA guidelines recommend a 6 week antibiotic therapy.[23] This is mainly based on a randomized clinical trial that showed that 6 weeks of antibiotic treatment is noninferior to 12 weeks. The 6-week recommendation is, also, supported by another retrospective study in which the first group was treated for less than 6 weeks and the second for more than 6 weeks. The outcomes, rates of relapse and deaths were comparable between the two groups.[84]

Treatment can be discontinued after 6 weeks in most patients with clinical improvement. However, those diagnosed with *Brucella*, Tuberculous or fungal infection should continue their therapy for the targeted duration.[4],[98] In case of complications such as abscess formation, the duration of treatment is prolonged.[99] Pediatric patients should receive intravenous antibiotics for about two weeks, followed by oral antibiotic for another one to three weeks if there is clinical and laboratory improvement.[99]

There is controversy regarding the switch from parenteral drug administration to oral. Intravenous antibiotics are used initially for 2 to 4 weeks in most cases. [30], [100] Recent studies argue that an early switch to agents with great oral bioavailability has similar efficacy to prolonged intravenous drug administration. [62],[101]

Discontinuation of antimicrobial therapy is considered in neurological deterioration with imaging tests indicating progressive destruction. Furthermore, a different approach should be considered if the expected clinical improvement is not achieved.[100] In either case, attempts to isolate a pathogen should be made.

9. Surgical management

A surgical approach is deemed necessary in case of failure of conservative measures.[102] Other indications for surgery are symptoms persistence, onset or progression of neurologic deficits, spinal instability, abscess larger than 2.5 cm, signs of ischemia or compression and deformities such as kyphosis or scoliosis. [103],[104] Urgent operation is indicated in septicemia

TABLE 3.

Criteria for absolute and relative surgery indications. (Saeed et al., 2019)

Indication for surgery	Absolute	Relative
Neurologic deficit	+	-
Spinal instability/ deformities (e.g. Kyphosis)	+	-
Spinal core compression/ cauda equina	With neurologic deficit	Without neurologic deficit
Space occupying/ non drainable abscess	+	-
Sepsis	+	-
Conservative treatment failure	-	+
Extensive spread of the infection	Antibiotics non responsive, clinical, laboratory, imaging deterioration with positive cultures	Without laboratory and clinical deterioration

or rapid clinical deterioration with no response to drug treatment.[30],[99]

Thorough surgical debridement and maintenance or restoration of vertebral stability are the principal goals. Open surgery with extensive debridement of the infected tissue is most times recommended while minimally invasive surgery is an alternative method. [105]


Anterior approach is indicated for anterior debridement and stabilization, whereas the posterior approach is indicated for decompression of a primary posterior epidural abscess with concomitant posterior spinal instrumentation.[106] A combined anterior-posterior approach has been occasionally used.[105],[107]

Thorough debridement may result in extensive tissue loss endangering the vertebral column's integrity. Therefore, instrumentation and bone grafting are used to stabilize the spine. However, some authors believe that metallic implants are possible foci for bacterial adherence.[103] Nevertheless, spinal instrumentation provides stability and increased fusion rates.[107] Moreover titanium alloy implants are less prone to colonization than stainless steel ones. [108] Additionally, less time of patient immobilization is required. [109]

In postoperative spinal infections with metallic im-

plant involvement, implant removal is most times mandatory.[67] However, stable grafts adherent to native bone should be left in place. If implant removal results in fracture of the fusion mass, bone grafting should be done to ensure alignment of the vertebral column.[110]

10. Conclusion

Spinal infection is a well-documented disease which predominantly affects people with certain risk factors and people from endemic areas. The most common pathogens are bacteria, especially *Staphylococcus* species. Diagnosis is quite challenging, requiring collaboration of physicians from different fields of medicine. Appropriate management remains an area of controversy. Most evidence-based guidelines along with experts' opinion recommend a conservative approach of antimicrobial drugs and patient immobilization. Surgical treatment may be considered in infection persistence, and extensive disease. Surgery involves broad debridement, bone grafting and spinal stabilization. Publication of more studies is crucial to ensure optimal diagnostic evaluation and disease management. 

Conflicts of Interest

The authors declare that they have no conflicts of Interest.

REFERENCES

- [1] D. M. Kaufman, J. G. Kaplan, and N. Litman, "Infectious agents in spinal epidural abscesses," *Neurology*, vol. 30, no. 8, pp. 844–850, Aug. 1980, doi: 10.1212/wnl.30.8.844.
- [2] R. R. Calderone and J. M. Larsen, "Overview and classification of spinal infections," *Orthop. Clin. North Am.*, vol. 27, no. 1, pp. 1–8, Jan. 1996.
- [3] L. Grammatico et al., "Epidemiology of vertebral osteomyelitis (VO) in France: analysis of hospital-discharge data 2002–2003," *Epidemiol. Infect.*, vol. 136, no. 5, pp. 653–660, May 2008, doi: 10.1017/S0950268807008850.
- [4] K. Y. Lee, "Comparison of Pyogenic Spondylitis and Tuberculous Spondylitis," *Asian Spine J.*, vol. 8, no. 2, p. 216, 2014, doi: 10.4184/asj.2014.8.2.216.
- [5] A. G. Jensen, F. Espersen, P. Skinhøj, V. T. Rosdahl, and N. Frimodt-Møller, "Increasing frequency of vertebral osteomyelitis following *Staphylococcus aureus* bacteraemia in Denmark 1980–1990," *J. Infect.*, vol. 34, no. 2, pp. 113–118, Mar. 1997, doi: 10.1016/S0163-4453(97)92395-1.
- [6] J. Solera, E. Lozano, E. Martinez-Alfaro, A. Espinosa, M. L. Castillejos, and L. Abad, "Brucellar Spondylitis: Review of 35 Cases and Literature Survey," *Clin. Infect. Dis.*, vol. 29, no. 6, pp. 1440–1449, Dec. 1999, doi: 10.1086/313524.
- [7] J. L. Cebrián Parra, A. Saez-Arenillas Martín, A. L. Urda Martínez-Aedo, I. Soler Ivañez, E. Agreda, and L. Lopez-Duran Stern, "Management of infectious discitis. Outcome in one hundred and eight patients in a University Hospital," *Int. Orthop.*, vol. 36, no. 2, pp. 239–244, Feb. 2012, doi: 10.1007/s00264-011-1445-x.
- [8] E. T. Tali, "Spinal infections," *Eur. J. Radiol.*, vol. 50, no. 2, pp. 120–133, May 2004, doi: 10.1016/j.ejrad.2003.10.022.
- [9] A. S. Smith and S. I. Blaser, "Infectious and inflammatory processes of the spine," *Radiol. Clin. North Am.*, vol. 29, no. 4, pp. 809–827, Jul. 1991.
- [10] M. J. Patzakis, S. Rao, J. Wilkins, T. M. Moore, and P. J. Harvey, "Analysis of 61 cases of vertebral osteomyelitis," *Clin. Orthop.*, no. 264, pp. 178–183, Mar. 1991.
- [11] J. D. Colmenero et al., "Pyogenic, tuberculous, and brucellar vertebral osteomyelitis: a descriptive and comparative study of 219 cases," *Ann. Rheum. Dis.*, vol. 56, no. 12, pp. 709–715, Dec. 1997, doi: 10.1136/ard.56.12.709.
- [12] J. F. Griffith, S. M. Kumta, P. C. Leung, J. C. Y. Cheng, L. T. C. Chow, and C. Metreweli, "Imaging of musculoskeletal tuberculosis: a new look at an old disease," *Clin. Orthop.*, no. 398, pp. 32–39, May 2002, doi: 10.1097/00003086-200205000-00006.
- [13] D. W. Park et al., "Outcome and management of spinal tuberculosis according to the severity of disease: a retrospective study of 137 adult patients at Korean teaching hospitals," *Spine*, vol. 32, no. 4, pp. E130–135, Feb. 2007, doi: 10.1097/01.brs.0000255216.54085.21.
- [14] D. L. Larson, K. A. Hudak, W. P. Waring, M. R. Orr, and K. Simonelic, "Protocol management of late-stage pressure ulcers: a 5-year retrospective study of 101 consecutive patients with 179 ulcers," *Plast. Reconstr. Surg.*, vol. 129, no. 4, pp. 897–904, Apr. 2012, doi: 10.1097/PRS.0b013e3182442197.
- [15] M. A. Ameer, T. L. Knorr, and F. B. Mesfin, "Spinal Epidural Abscess," in *StatPearls*, Treasure Island (FL): StatPearls Publishing, 2020.
- [16] C. Michel-Batôt et al., "A particular form of septic arthritis: septic arthritis of facet joint," *Joint Bone Spine*, vol. 75, no. 1, pp. 78–83, Jan. 2008, doi: 10.1016/j.jb-spine.2007.02.006.
- [17] V. Dufour et al., "Comparative study of postoperative and spontaneous pyogenic spondylodiscitis," *Semin. Arthritis Rheum.*, vol. 34, no. 5, pp. 766–771, Apr. 2005, doi: 10.1016/j.semarthrit.2004.08.004.
- [18] M. A. Weinstein and F. J. Eismont, "Infections of the spine in patients with human immunodeficiency virus," *J. Bone Joint Surg. Am.*, vol. 87, no. 3, pp. 604–609, Mar. 2005, doi: 10.2106/JBJS.C.01062.
- [19] A. Di Martino, R. Papalia, E. Albo, L. Diaz, L. Denaro, and V. Denaro, "Infection after spinal surgery and procedures," *Eur. Rev. Med. Pharmacol. Sci.*, vol. 23, no. 2 Suppl, pp. 173–178, Apr. 2019, doi: 10.26355/eu-rrev_201904_17487.
- [20] N. A. S. Sai Kiran, S. Vaishya, S. S. Kale, B. S. Sharma, and A. K. Mahapatra, "Surgical results in patients

- with tuberculosis of the spine and severe lower-extremity motor deficits: a retrospective study of 48 patients," *J. Neurosurg. Spine*, vol. 6, no. 4, pp. 320–326, Apr. 2007, doi: 10.3171/spi.2007.6.4.6.
- [21] A. S. Baker, R. G. Ojemann, M. N. Swartz, and E. P. Richardson, "Spinal epidural abscess," *N. Engl. J. Med.*, vol. 293, no. 10, pp. 463–468, Sep. 1975, doi: 10.1056/NEJM197509042931001.
- [22] A. M. Wiley and J. Trueta, "The vascular anatomy of the spine and its relationship to pyogenic vertebral osteomyelitis," *J. Bone Joint Surg. Br.*, vol. 41-B, pp. 796–809, Nov. 1959, doi: 10.1302/0301-620X.41B4.796.
- [23] E. F. Berbari et al., "2015 Infectious Diseases Society of America (IDSA) Clinical Practice Guidelines for the Diagnosis and Treatment of Native Vertebral Osteomyelitis in Adults," *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.*, vol. 61, no. 6, pp. e26–46, Sep. 2015, doi: 10.1093/cid/civ482.
- [24] A. Rivas-Garcia, S. Sarria-Estrada, C. Torrents-Odin, L. Casas-Gomila, and E. Franquet, "Imaging findings of Pott's disease," *Eur. Spine J. Off. Publ. Eur. Spine Soc. Eur. Spinal Deform. Soc. Eur. Sect. Cerv. Spine Res. Soc.*, vol. 22 Suppl 4, pp. 567–578, Jun. 2013, doi: 10.1007/s00586-012-2333-9.
- [25] M. N. Gamaletsou et al., "Aspergillus osteomyelitis: epidemiology, clinical manifestations, management, and outcome," *J. Infect.*, vol. 68, no. 5, pp. 478–493, May 2014, doi: 10.1016/j.jinf.2013.12.008.
- [26] P. Kapeller et al., "Pyogenic infectious spondylitis: clinical, laboratory and MRI features," *Eur. Neurol.*, vol. 38, no. 2, pp. 94–98, 1997, doi: 10.1159/000113167.
- [27] A. F. Mavrogenis, G. K. Triantafyllopoulos, K. Kokkinis, A. Stefanos, N. V. Sipsas, and S. G. Pneumatikos, "Continuous L3 spondylitis caused by an infected endovascular aortic graft," *Surg. Infect.*, vol. 15, no. 6, pp. 861–862, Dec. 2014, doi: 10.1089/sur.2013.219.
- [28] O. V. Batson, "The vertebral system of veins as a means for cancer dissemination," *Prog. Clin. Cancer*, vol. 3, pp. 1–18, 1967.
- [29] W.-C. Chang, H.-K. Tsou, T.-H. Kao, M.-Y. Yang, and C.-C. Shen, "Successful treatment of extended epidural abscess and long segment osteomyelitis: a case report and review of the literature," *Surg. Neurol.*, vol. 69, no. 2, pp. 117–120; discussion 120, Feb. 2008, doi: 10.1016/j.surneu.2006.12.047.
- [30] F. L. Sapico and J. Z. Montgomerie, "Pyogenic vertebral osteomyelitis: report of nine cases and review of the literature," *Rev. Infect. Dis.*, vol. 1, no. 5, pp. 754–776, Oct. 1979, doi: 10.1093/clinids/1.5.754.
- [31] M. Fantoni et al., "Epidemiological and clinical features of pyogenic spondylodiscitis," *Eur. Rev. Med. Pharmacol. Sci.*, vol. 16 Suppl 2, pp. 2–7, Apr. 2012.
- [32] J. S. Butler, M. J. Shelly, M. Timlin, W. G. Powderly, and J. M. O'Byrne, "Nontuberculous pyogenic spinal infection in adults: a 12-year experience from a tertiary referral center," *Spine*, vol. 31, no. 23, pp. 2695–2700, Nov. 2006, doi: 10.1097/01.brs.0000244662.78725.37.
- [33] C. E. C. Goertz et al., "Brucella sp. vertebral osteomyelitis with intercurrent fatal Staphylococcus aureus toxigenic enteritis in a bottlenose dolphin (*Tursiops truncatus*)," *J. Vet. Diagn. Investig. Off. Publ. Am. Assoc. Vet. Lab. Diagn. Inc.*, vol. 23, no. 4, pp. 845–851, Jul. 2011, doi: 10.1177/1040638711407683.
- [34] A. L. Gasbarrini et al., "Clinical features, diagnostic and therapeutic approaches to haematogenous vertebral osteomyelitis," *Eur. Rev. Med. Pharmacol. Sci.*, vol. 9, no. 1, pp. 53–66, Feb. 2005.
- [35] W. T. Davis, M. D. April, S. Mehta, B. Long, and S. Shroyer, "High risk clinical characteristics for pyogenic spinal infection in acute neck or back pain: Prospective cohort study," *Am. J. Emerg. Med.*, vol. 38, no. 3, pp. 491–496, 2020, doi: 10.1016/j.ajem.2019.05.025.
- [36] W. Y. Cheung and K. D. K. Luk, "Pyogenic spondylitis," *Int. Orthop.*, vol. 36, no. 2, pp. 397–404, Feb. 2012, doi: 10.1007/s00264-011-1384-6.
- [37] A. F. Mavrogenis et al., "Spondylodiscitis revisited," *EFORT Open Rev.*, vol. 2, no. 11, pp. 447–461, Nov. 2017, doi: 10.1302/2058-5241.2.160062.
- [38] T. Aagaard, C. Roed, C. Dragsted, and P. Skinhøj, "Microbiological and therapeutic challenges in infectious spondylodiscitis: a cohort study of 100 cases, 2006–2011," *Scand. J. Infect. Dis.*, vol. 45, no. 6, pp. 417–424, Jun. 2013, doi: 10.3109/00365548.2012.753160.
- [39] S. Choi et al., "Diagnostic usefulness of the QuantiFERON-TB gold in-tube test (QFT-GIT) for tuberculous vertebral osteomyelitis," *Infect. Dis.*

- Lond. Engl., vol. 50, no. 5, pp. 346–351, 2018, doi: 10.1080/23744235.2017.1410282.
- [40] E. Mylona, M. Samarkos, E. Kakalou, P. Fanourgiakis, and A. Skoutelis, "Pyogenic vertebral osteomyelitis: a systematic review of clinical characteristics," *Semin. Arthritis Rheum.*, vol. 39, no. 1, pp. 10–17, Aug. 2009, doi: 10.1016/j.semarthrit.2008.03.002.
- [41] S. Rajasekaran, "The problem of deformity in spinal tuberculosis," *Clin. Orthop.*, no. 398, pp. 85–92, May 2002, doi: 10.1097/00003086-200205000-00012.
- [42] V. Jevtic, "Vertebral infection," *Eur. Radiol.*, vol. 14 Suppl 3, pp. E43–52, Mar. 2004, doi: 10.1007/s00330-003-2046-x.
- [43] A. Dagirmanjian, J. Schils, and M. C. McHenry, "MR imaging of spinal infections," *Magn. Reson. Imaging Clin. N. Am.*, vol. 7, no. 3, pp. 525–538, Aug. 1999.
- [44] F. Maiuri, G. Iaconetta, B. Gallicchio, A. Manto, and F. Briganti, "Spondylodiscitis. Clinical and magnetic resonance diagnosis," *Spine*, vol. 22, no. 15, pp. 1741–1746, Aug. 1997, doi: 10.1097/00007632-199708010-00012.
- [45] M. M. Thurnher and R. Bammer, "Diffusion-weighted magnetic resonance imaging of the spine and spinal cord," *Semin. Roentgenol.*, vol. 41, no. 4, pp. 294–311, Oct. 2006, doi: 10.1053/j.ro.2006.07.003.
- [46] E. J. Carragee, "Pyogenic vertebral osteomyelitis," *J. Bone Joint Surg. Am.*, vol. 79, no. 6, pp. 874–880, Jun. 1997, doi: 10.2106/00004623-199706000-00011.
- [47] E. Lazzeri et al., "Clinical feasibility of two-step streptavidin/111In-biotin scintigraphy in patients with suspected vertebral osteomyelitis," *Eur. J. Nucl. Med. Mol. Imaging*, vol. 31, no. 11, pp. 1505–1511, Nov. 2004, doi: 10.1007/s00259-004-1581-2.
- [48] A. Lupetti, M. M. Welling, U. Mazzi, P. H. Nibbering, and E. K. J. Pauwels, "Technetium-99m labelled fluconazole and antimicrobial peptides for imaging of *Candida albicans* and *Aspergillus fumigatus* infections," *Eur. J. Nucl. Med. Mol. Imaging*, vol. 29, no. 5, pp. 674–679, May 2002, doi: 10.1007/s00259-001-0760-7.
- [49] A. S. Tamm and J. T. Abele, "Bone and Gallium Single-Photon Emission Computed Tomography-Computed Tomography is Equivalent to Magnetic Resonance Imaging in the Diagnosis of Infectious Spondylodiscitis: A Retrospective Study," *Can. Assoc. Radiol. J. J. Assoc. Can. Radiol.*, vol. 68, no. 1, pp. 41–46, Feb. 2017, doi: 10.1016/j.carj.2016.02.003.
- [50] S. Gratz et al., "18F-FDG hybrid PET in patients with suspected spondylitis," *Eur. J. Nucl. Med. Mol. Imaging*, vol. 29, no. 4, pp. 516–524, Apr. 2002, doi: 10.1007/s00259-001-0719-8.
- [51] K. Strobel and K. D. M. Stumpe, "PET/CT in musculoskeletal infection," *Semin. Musculoskelet. Radiol.*, vol. 11, no. 4, pp. 353–364, Dec. 2007, doi: 10.1055/s-2008-1060337.
- [52] S. C. Foreman et al., "MR and CT Imaging to Optimize CT-Guided Biopsies in Suspected Spondylodiscitis," *World Neurosurg.*, vol. 99, pp. 726–734.e7, Mar. 2017, doi: 10.1016/j.wneu.2016.11.017.
- [53] F. S. Chew and M. J. Kline, "Diagnostic yield of CT-guided percutaneous aspiration procedures in suspected spontaneous infectious diskitis," *Radiology*, vol. 218, no. 1, pp. 211–214, Jan. 2001, doi: 10.1148/radiology.218.1.r01ja06211.
- [54] A. Olscamp, J. Rollins, S. S. Tao, and N. A. Ebraheim, "Complications of CT-guided biopsy of the spine and sacrum," *Orthopedics*, vol. 20, no. 12, pp. 1149–1152, Dec. 1997.
- [55] F. Lecouvet, L. Irenge, B. Vandercam, A. Nzeusseu, S. Hamels, and J.-L. Gala, "The etiologic diagnosis of infectious discitis is improved by amplification-based DNA analysis," *Arthritis Rheum.*, vol. 50, no. 9, pp. 2985–2994, Sep. 2004, doi: 10.1002/art.20462.
- [56] G. Wang et al., "Diagnostic accuracy evaluation of the conventional and molecular tests for Spinal Tuberculosis in a cohort, head-to-head study," *Emerg. Microbes Infect.*, vol. 7, no. 1, p. 109, Jun. 2018, doi: 10.1038/s41426-018-0114-1.
- [57] G. Gras et al., "Microbiological diagnosis of vertebral osteomyelitis: relevance of second percutaneous biopsy following initial negative biopsy and limited yield of post-biopsy blood cultures," *Eur. J. Clin. Microbiol. Infect. Dis. Off. Publ. Eur. Soc. Clin. Microbiol.*, vol. 33, no. 3, pp. 371–375, Mar. 2014, doi: 10.1007/s10096-013-1965-y.
- [58] K. Fuursted, M. Arpi, B. E. Lindblad, and L. N. Ped-

- ersen, "Broad-range PCR as a supplement to culture for detection of bacterial pathogens in patients with a clinically diagnosed spinal infection," *Scand. J. Infect. Dis.*, vol. 40, no. 10, pp. 772–777, 2008, doi: 10.1080/00365540802119994.
- [59] A. Navarro-Martínez, E. Navarro, M. J. Castaño, and J. Solera, "Rapid diagnosis of human brucellosis by quantitative real-time PCR: a case report of brucellar spondylitis," *J. Clin. Microbiol.*, vol. 46, no. 1, pp. 385–387, Jan. 2008, doi: 10.1128/JCM.01303-07.
- [60] E. M. de Lucas et al., "CT-guided fine-needle aspiration in vertebral osteomyelitis: true usefulness of a common practice," *Clin. Rheumatol.*, vol. 28, no. 3, pp. 315–320, Mar. 2009, doi: 10.1007/s10067-008-1051-5.
- [61] J. Marschall, K. P. Bhavan, M. A. Olsen, V. J. Fraser, N. M. Wright, and D. K. Warren, "The impact of prebiopsy antibiotics on pathogen recovery in hematogenous vertebral osteomyelitis," *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.*, vol. 52, no. 7, pp. 867–872, Apr. 2011, doi: 10.1093/cid/cir062.
- [62] W. Zimmerli, "Clinical practice. Vertebral osteomyelitis," *N. Engl. J. Med.*, vol. 362, no. 11, pp. 1022–1029, Mar. 2010, doi: 10.1056/NEJMcp0910753.
- [63] F. Grados, F. X. Lescure, E. Senneville, R. M. Flipo, J. L. Schmit, and P. Fardellone, "Suggestions for managing pyogenic (non-tuberculous) discitis in adults," *Joint Bone Spine*, vol. 74, no. 2, pp. 133–139, Mar. 2007, doi: 10.1016/j.jbspin.2006.11.002.
- [64] D. K. H. Yee, D. Samartzis, Y.-W. Wong, K. D. K. Luk, and K. M. C. Cheung, "Infective spondylitis in Southern Chinese: a descriptive and comparative study of ninety-one cases," *Spine*, vol. 35, no. 6, pp. 635–641, Mar. 2010, doi: 10.1097/BRS.0b013e3181cf4f6.
- [65] R. K. Garg and D. S. Somvanshi, "Spinal tuberculosis: a review," *J. Spinal Cord Med.*, vol. 34, no. 5, pp. 440–454, 2011, doi: 10.1179/2045772311Y.0000000023.
- [66] C. B. Josephson, S. Al-Azri, D. J. Smyth, D. Haase, and B. L. Johnston, "A case of Pott's disease with epidural abscess and probable cerebral tuberculoma following Bacillus Calmette-Guérin therapy for superficial bladder cancer," *Can. J. Infect. Dis. Med. Microbiol. J. Can. Mal. Infect. Microbiol. Med. J. Can.*, vol. 21, no. 1, pp. e75–78, 2010, doi: 10.1155/2010/572410.
- [67] M. Loibl et al., "Outcome-related co-factors in 105 cases of vertebral osteomyelitis in a tertiary care hospital," *Infection*, vol. 42, no. 3, pp. 503–510, Jun. 2014, doi: 10.1007/s15010-013-0582-0.
- [68] J. M. Greig and M. J. Wood, "Staphylococcus lugdunensis vertebral osteomyelitis," *Clin. Microbiol. Infect. Off. Publ. Eur. Soc. Clin. Microbiol. Infect. Dis.*, vol. 9, no. 11, pp. 1139–1141, Nov. 2003, doi: 10.1046/j.1469-0691.2003.00777.x.
- [69] L. Cottle and T. Riordan, "Infectious spondylodiscitis," *J. Infect.*, vol. 56, no. 6, pp. 401–412, Jun. 2008, doi: 10.1016/j.jinf.2008.02.005.
- [70] F. A. Broner, D. E. Garland, and J. E. Zigler, "Spinal infections in the immunocompromised host," *Orthop. Clin. North Am.*, vol. 27, no. 1, pp. 37–46, Jan. 1996.
- [71] R. Tyagi, "Spinal infections in children: A review," *J. Orthop.*, vol. 13, no. 4, pp. 254–258, Dec. 2016, doi: 10.1016/j.jor.2016.06.005.
- [72] C. Hopf, A. Meurer, P. Eysel, and J. D. Rompe, "Operative treatment of spondylodiscitis—what is the most effective approach?," *Neurosurg. Rev.*, vol. 21, no. 4, pp. 217–225, 1998, doi: 10.1007/BF01105775.
- [73] C. Schinkel, M. Gottwald, and H.-J. Andress, "Surgical treatment of spondylodiscitis," *Surg. Infect.*, vol. 4, no. 4, pp. 387–391, 2003, doi: 10.1089/109629603322761445.
- [74] C.-Y. Chuo et al., "Spinal infection in intravenous drug abusers," *J. Spinal Disord. Tech.*, vol. 20, no. 4, pp. 324–328, Jun. 2007, doi: 10.1097/BSD.0b013e-31802c144a.
- [75] Z. Chen, P. Cao, Z. Zhou, Y. Yuan, Y. Jiao, and Y. Zheng, "Overview: the role of Propionibacterium acnes in nonpyogenic intervertebral discs," *Int. Orthop.*, vol. 40, no. 6, pp. 1291–1298, Jun. 2016, doi: 10.1007/s00264-016-3115-5.
- [76] K. Saeed et al., "Hot topics on vertebral osteomyelitis from the International Society of Antimicrobial Chemotherapy," *Int. J. Antimicrob. Agents*, vol. 54, no. 2, pp. 125–133, Aug. 2019, doi: 10.1016/j.ijantimicag.2019.06.013.
- [77] E. F. Berbari et al., "2015 Infectious Diseases Society of America (IDSA) Clinical Practice Guidelines for the Diagnosis and Treatment of Native Vertebral Osteomyelitis in Adults," *Clin. Infect. Dis.*, vol. 61, no. 6, pp. e26–e46, Sep. 2015, doi: 10.1093/cid/civ482.
- [78] E. Mylona, M. Samarkos, E. Kakalou, P. Fanourgiakis,

- and A. Skoutelis, "Pyogenic Vertebral Osteomyelitis: A Systematic Review of Clinical Characteristics," *Semin. Arthritis Rheum.*, vol. 39, no. 1, pp. 10–17, Aug. 2009, doi: 10.1016/j.semarthrit.2008.03.002.
- [79] Z. A. Memish and H. H. Balkhy, "Brucellosis and international travel," *J. Travel Med.*, vol. 11, no. 1, pp. 49–55, Feb. 2004, doi: 10.2310/7060.2004.13551.
- [80] L. I. Sakkas et al., "Hematogenous spinal infection in central Greece," *Spine*, vol. 34, no. 15, pp. E513–518, Jul. 2009, doi: 10.1097/BRS.0b013e3181a9897e.
- [81] L. D. Herron, P. Kissel, and D. Smilovitz, "Treatment of coccidioidal spinal infection: experience in 16 cases," *J. Spinal Disord.*, vol. 10, no. 3, pp. 215–222, Jun. 1997.
- [82] T. N. Joshi, "Candida albicans spondylodiscitis in an immunocompetent patient," *J. Neurosci. Rural Pract.*, vol. 3, no. 2, pp. 221–222, May 2012, doi: 10.4103/0976-3147.98261.
- [83] A. Gennari, F. Almairac, S. Litrico, C. Albert, P. Marty, and P. Paquis, "Spinal cord compression due to a primary vertebral hydatid disease: A rare case report in metropolitan France and a literature review," *Neurochirurgie*, vol. 62, no. 4, pp. 226–228, Aug. 2016, doi: 10.1016/j.neuchi.2016.03.001.
- [84] F. Roblot et al., "Optimal duration of antibiotic therapy in vertebral osteomyelitis," *Semin. Arthritis Rheum.*, vol. 36, no. 5, pp. 269–277, Apr. 2007, doi: 10.1016/j.semarthrit.2006.09.004.
- [85] E. Pola et al., "New classification for the treatment of pyogenic spondylodiscitis: validation study on a population of 250 patients with a follow-up of 2 years," *Eur. Spine J. Off. Publ. Eur. Spine Soc. Eur. Spinal Deform. Soc. Eur. Sect. Cerv. Spine Res. Soc.*, vol. 26, no. Suppl 4, pp. 479–488, 2017, doi: 10.1007/s00586-017-5043-5.
- [86] P. Viale et al., "Treatment of pyogenic (non-tuberculous) spondylodiscitis with tailored high-dose levofloxacin plus rifampicin," *Int. J. Antimicrob. Agents*, vol. 33, no. 4, pp. 379–382, Apr. 2009, doi: 10.1016/j.ijantimicag.2008.09.011.
- [87] C. Liu et al., "Clinical Practice Guidelines by the Infectious Diseases Society of America for the Treatment of Methicillin-Resistant *Staphylococcus aureus* Infections in Adults and Children: Executive Summary," *Clin. Infect. Dis.*, vol. 52, no. 3, pp. 285–292, Feb. 2011, doi: 10.1093/cid/cir034.
- [88] Baddour Larry M. et al., "Infective Endocarditis," *Circulation*, vol. 111, no. 23, pp. e394–e434, Jun. 2005, doi: 10.1161/CIRCULATIONAHA.105.165564.
- [89] W. Graninger and R. Raggette, "Nosocomial Bacteremia Due to *Enterococcus faecalis* without Endocarditis," *Clin. Infect. Dis.*, vol. 15, no. 1, pp. 49–57, Jul. 1992, doi: 10.1093/clinids/15.1.49.
- [90] A. D. Tice et al., "Practice Guidelines for Outpatient Parenteral Antimicrobial Therapy," *Clin. Infect. Dis.*, vol. 38, no. 12, pp. 1651–1671, Jun. 2004, doi: 10.1086/420939.
- [91] H. M. Blumberg et al., "American Thoracic Society/ Centers for Disease Control and Prevention/ Infectious Diseases Society of America: treatment of tuberculosis," *Am. J. Respir. Crit. Care Med.*, vol. 167, no. 4, pp. 603–662, Feb. 2003, doi: 10.1164/rccm.167.4.603.
- [92] T. Shi et al., "Retrospective Study of 967 Patients With Spinal Tuberculosis," *Orthopedics*, vol. 39, no. 5, pp. e838–843, Sep. 2016, doi: 10.3928/01477447-20160509-03.
- [93] S. W. Chapman et al., "Clinical practice guidelines for the management of blastomycosis: 2008 update by the Infectious Diseases Society of America," *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.*, vol. 46, no. 12, pp. 1801–1812, Jun. 2008, doi: 10.1086/588300.
- [94] T. J. Walsh et al., "Treatment of aspergillosis: clinical practice guidelines of the Infectious Diseases Society of America," *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.*, vol. 46, no. 3, pp. 327–360, Feb. 2008, doi: 10.1086/525258.
- [95] P. J. Carek, L. M. Dickerson, and J. L. Sack, "Diagnosis and management of osteomyelitis," *Am. Fam. Physician*, vol. 63, no. 12, pp. 2413–2420, Jun. 2001.
- [96] L. Lazzarini, F. De Lalla, and J. T. Mader, "Long Bone Osteomyelitis," *Curr. Infect. Dis. Rep.*, vol. 4, no. 5, pp. 439–445, Oct. 2002, doi: 10.1007/s11908-002-0012-4.
- [97] N. Bettini, M. Girardo, E. Dema, and S. Cervellati, "Evaluation of conservative treatment of non specific spondylodiscitis," *Eur. Spine J. Off. Publ. Eur. Spine Soc. Eur. Spinal Deform. Soc. Eur. Sect. Cerv. Spine*

- Res. Soc., vol. 18 Suppl 1, pp. 143–150, Jun. 2009, doi: 10.1007/s00586-009-0979-8.
- [98] M. Chelli Bouaziz, M. F. Ladeb, M. Chakroun, and S. Chaabane, "Spinal brucellosis: a review," *Skeletal Radiol.*, vol. 37, no. 9, pp. 785–790, Sep. 2008, doi: 10.1007/s00256-007-0371-x.
- [99] C. Liu et al., "Clinical practice guidelines by the infectious diseases society of america for the treatment of methicillin-resistant *Staphylococcus aureus* infections in adults and children: executive summary," *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.*, vol. 52, no. 3, pp. 285–292, Feb. 2011, doi: 10.1093/cid/cir034.
- [100] K. Zarghooni, M. Rölinghoff, R. Sobottke, and P. Eysel, "Treatment of spondylodiscitis," *Int. Orthop.*, vol. 36, no. 2, pp. 405–411, Feb. 2012, doi: 10.1007/s00264-011-1425-1.
- [101] H.-K. Li et al., "Oral versus Intravenous Antibiotics for Bone and Joint Infection," *N. Engl. J. Med.*, vol. 380, no. 5, pp. 425–436, 31 2019, doi: 10.1056/NEJ-Moa1710926.
- [102] P. C. Hsieh, R. J. Wienecke, B. A. O'Shaughnessy, T. R. Koski, and S. L. Ondra, "Surgical strategies for vertebral osteomyelitis and epidural abscess," *Neurosurg. Focus*, vol. 17, no. 6, p. E4, Dec. 2004, doi: 10.3171/foc.2004.17.6.4.
- [103] W.-H. Chen, L.-S. Jiang, and L.-Y. Dai, "Surgical treatment of pyogenic vertebral osteomyelitis with spinal instrumentation," *Eur. Spine J. Off. Publ. Eur. Spine Soc. Eur. Spinal Deform. Soc. Eur. Sect. Cerv. Spine Res. Soc.*, vol. 16, no. 9, pp. 1307–1316, Sep. 2007, doi: 10.1007/s00586-006-0251-4.
- [104] L. A. Nasto et al., "Is posterior percutaneous screw-rod instrumentation a safe and effective alternative approach to TLSO rigid bracing for single-level pyogenic spondylodiscitis? Results of a retrospective cohort analysis," *Spine J. Off. J. North Am. Spine Soc.*, vol. 14, no. 7, pp. 1139–1146, Jul. 2014, doi: 10.1016/j.spinee.2013.07.479.
- [105] P. Korovessis, V. Syrimpeis, V. Tsekouras, A. Baikoussis, K. Vardakastanis, and P. Fennema, "A unilateral less invasive posterolateral approach for disc debridement and titanium cage insertion supplemented by contralateral transfascial screw fixation for high-morbidity patients suffering from septic thoracolumbosacral spondylodiscitis," *Eur. J. Orthop. Surg. Traumatol. Orthop. Traumatol.*, vol. 29, no. 6, pp. 1187–1197, Aug. 2019, doi: 10.1007/s00590-019-02434-2.
- [106] S. A. Rath, U. Neff, O. Schneider, and H. P. Richter, "Neurosurgical management of thoracic and lumbar vertebral osteomyelitis and discitis in adults: a review of 43 consecutive surgically treated patients," *Neurosurgery*, vol. 38, no. 5, pp. 926–933, May 1996, doi: 10.1097/00006123-199605000-00013.
- [107] A. F. Mavrogenis et al., "When and how to operate on spondylodiscitis: a report of 13 patients," *Eur. J. Orthop. Surg. Traumatol. Orthop. Traumatol.*, vol. 26, no. 1, pp. 31–40, Jan. 2016, doi: 10.1007/s00590-015-1674-6.
- [108] G. D. Sundararaj, R. Amritanand, K. Venkatesh, and J. Arockiaraj, "The use of titanium mesh cages in the reconstruction of anterior column defects in active spinal infections: can we rest the crest?," *Asian Spine J.*, vol. 5, no. 3, pp. 155–161, Sep. 2011, doi: 10.4184/asj.2011.5.3.155.
- [109] E. J. Karadimas et al., "Spondylodiscitis. A retrospective study of 163 patients," *Acta Orthop.*, vol. 79, no. 5, pp. 650–659, Oct. 2008, doi: 10.1080/17453670810016678.
- [110] M. Di Silvestre, G. Bakaloudis, F. Lolli, and S. Giacomini, "Late-developing infection following posterior fusion for adolescent idiopathic scoliosis," *Eur. Spine J. Off. Publ. Eur. Spine Soc. Eur. Spinal Deform. Soc. Eur. Sect. Cerv. Spine Res. Soc.*, vol. 20 Suppl 1, pp. S121–127, May 2011, doi: 10.1007/s00586-011-1754-1.